

# Copper Ions Reduced Toxicity of Sodium Azide and Lipopolysaccharide on Cultured Cerebellar Granule Neurons

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## Abstract

**Introduction.** Copper ions ( $\text{Cu}^{2+}$ ) are structural elements of proteins such as cytochrome c oxidase (Complex IV), an enzyme that catalyzes the final step of electron transfer to oxygen during oxidative phosphorylation in the mitochondria. With  $\text{Cu}^{2+}$  homeostasis being of utmost importance, its disturbances in the central nervous system are involved in the mechanisms of many neurodegenerative and other brain disorders.

**This study aimed** to assess the effects of non-toxic copper ion levels on death of cultured cerebellar granule neurons associated with lipopolysaccharide (LPS; *in vitro* inflammation model) or azide sodium ( $\text{NaN}_3$ ; cytochrome c oxidase inhibitor).

**Materials and methods.** LPS (10  $\mu\text{g}/\text{mL}$ ) or  $\text{NaN}_3$  (250  $\mu\text{M}$ ) was added on day 7 to 8 to the culture medium with rat cerebellar cells for 24 hours *in vitro*. Nitrite concentrations were measured in the culture medium by Griess assay; absorbance was recorded with a spectrophotometer at 540 nm, and morphologically intact cells were counted as survived neurons.

**Results.** Added to the culture medium, LPS or  $\text{NaN}_3$  reduced neuron survival to  $15 \pm 2\%$  or  $20 \pm 3\%$  vs. control, respectively.  $\text{Cu}^{2+}$  (0.5 to 5.0  $\mu\text{M}$ ) increased neuron survival in a dose-dependent manner to  $78 \pm 4\%$  with toxic levels of LPS and to  $86 \pm 6\%$  with  $\text{NaN}_3$  with 5  $\mu\text{M}$   $\text{Cu}^{2+}$ . The concentration of nitrites in the control culture medium was  $2.0 \pm 0.2 \mu\text{M}$ . Added to the cell cultures, LPS increased the concentration of nitrites to  $8.5 \pm 0.5 \mu\text{M}$ .  $\text{Cu}^{2+}$  5  $\mu\text{M}$  did not show any significant effects on nitrite accumulation in the culture medium.

**Conclusions.** We showed that copper ions can exert protective effects on neurons against LPS-induced or  $\text{NaN}_3$ -induced toxicity. This protection is likely to be associated rather with  $\text{Cu}^{2+}$  interaction with Complex IV of the electron transfer chain in the mitochondria than with inhibition of NO production. Effects of  $\text{Cu}^{2+}$  on apoptosis pathway proteins also cannot be ruled out.

**Keywords:** neurons; copper ions; sodium azide; nitrogen oxide

**Ethics approval.** Authors confirm compliance with institutional and national standards for the use of laboratory animals in accordance with «Consensus Author Guidelines for Animal Use» (IAVES, 23 July 2010). The research protocol was approved by the Local Ethics Committee of the Research Center of Neurology (protocol No. 5-5/22, June 1, 2022).

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**Conflict of interest.** The authors declare no apparent or potential conflicts of interest related to the publication of this article.

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# Ионы меди снижают токсическое действие азид натрия и липополисахарида на культивированные зернистые нейроны мозжечка

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## Аннотация

**Введение.** Ионы меди ( $\text{Cu}^{2+}$ ) являются структурными элементами белков, в том числе цитохром *c*-оксидазы (комплекс IV) — фермента, катализирующего конечный этап переноса электронов на кислород в процессе окислительного фосфорилирования в митохондриях. Поддержание гомеостаза  $\text{Cu}^{2+}$  в головном мозге очень важно, и его нарушение в центральной нервной системе вовлечено в патогенез многих нейродегенеративных заболеваний и патологических состояний головного мозга.

**Цель исследования** — определить влияние нетоксических концентраций ионов меди на гибель культивированных зернистых нейронов мозжечка, вызванную липополисахаридом (ЛПС; модель воспаления *in vitro*) и азидом натрия ( $\text{NaN}_3$ , ингибитор цитохром *c*-оксидазы).

**Материалы и методы.** ЛПС (10 мкг/мл) или  $\text{NaN}_3$  (250 мкМ) добавляли на 7–8-й день *in vitro* в среду культивирования клеток мозжечка крыс на 24 ч. Уровень нитритов измеряли в среде культивирования методом Грисса, оптическую плотность регистрировали при длине волны 540 нм с помощью спектрофотометра, а число живых нейронов оценивали методом подсчёта морфологически интактных клеток.

**Результаты.** Добавление в среду культивирования ЛПС снижало выживаемость нейронов до  $15 \pm 2\%$  относительно контроля, а  $\text{NaN}_3$  — до  $20 \pm 3\%$ . В присутствии  $\text{Cu}^{2+}$  (0,5–5,0 мкМ) выживаемость нейронов дозозависимо повышалась: на фоне 5 мкМ  $\text{Cu}^{2+}$  при токсическом воздействии ЛПС — до  $78 \pm 4\%$ , а при действии  $\text{NaN}_3$  — до  $86 \pm 6\%$ . В среде культивирования контрольных культур содержание нитритов составляло  $2,0 \pm 0,2$  мкМ. Добавление ЛПС вызывало повышение уровня нитритов до  $8,5 \pm 0,5$  мкМ. Ионы меди не оказывали достоверного влияния на накопление нитритов в среде культивирования.

**Заключение.** Показана возможность защитного действия ионов меди на нейроны при токсичности, вызванной ЛПС и  $\text{NaN}_3$ . Видимо, эта защита обусловлена взаимодействием  $\text{Cu}^{2+}$  с комплексом IV цепи переноса электронов в митохондриях, а не подавлением продукции оксида азота, не исключено также влияние  $\text{Cu}^{2+}$  на белки путей апоптоза.

**Ключевые слова:** нейроны; ионы меди; азид натрия; оксид азота

**Этическое утверждение.** Авторы подтверждают соблюдение институциональных и национальных стандартов по использованию лабораторных животных в соответствии с «Consensus Author Guidelines for Animal Use» (IAVES, 23.07.2010). Протокол исследования одобрен Локальным этическим комитетом ФГБНУ НЦН (протокол № 5-5/22 от 01.06.2022).

**Источник финансирования.** Авторы заявляют об отсутствии внешних источников финансирования при проведении исследования.

**Конфликт интересов.** Авторы декларируют отсутствие явных и потенциальных конфликтов интересов, связанных с публикацией настоящей статьи.

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## Introduction

Copper is one of the most abundant transition metals in the human body. It takes part in oxygen metabolism, collagen synthesis, and skin pigmentation, maintaining the integrity of blood vessels, as well as in iron homeostasis, antioxidant defense, and neurotransmitter synthesis [1].  $\text{Cu}^{2+}$  ions are structural elements of several proteins. For instance, copper is an essential component of cytochrome *c* oxidase (Complex IV), an enzyme that catalyzes the final step of electron

transfer to oxygen during oxidative phosphorylation in the mitochondria. Copper ions are also contained in the superoxide dismutase molecule, which is the most important antioxidant, and ceruloplasmin, a blood plasma protein involved in the mechanisms of pro-oxidant and antioxidant reactions. Copper is also necessary for several important processes in the brain tissue, such as the regulation of intracellular signal transduction, catecholamine balance, myelination of neuron axons, and synaptic transmission in the central nervous system (CNS) [2].

The copper content in the brain ranges from approximately 3 to 5  $\mu\text{g/g}$  wet weight [1]. Recommended copper intake to maintain systemic homeostasis in adults is 0.8 to 2.4 mg/day [3]. Stable  $\text{Cu}^{2+}$  homeostasis in the brain is essential, and its disturbances can be fatal for neurons.  $\text{Cu}^{2+}$  homeostasis disorders in the CNS are involved in the mechanisms of many neurodegenerative and other brain disorders, such as Wilson disease and Alzheimer's disease [4–6].

Intracellular copper and iron imbalance can increase free radical production and oxidative stress [78] because these transition metals directly participate in the Fenton reaction, which results in hydroxyl radicals with high toxicity [9]. Two-valent copper can mediate generation of hydrogen peroxide by tau-protein [9] and increase effects of pro-oxidants. Micromolar concentrations of the antioxidant acetylcysteine in the culture medium showed pro-oxidant activity with nanomolar concentrations of copper [10]. However, very limited literature data are available on direct effects of these ions on key neurodegeneration processes, including inflammatory processes in the CNS and mitochondria inhibition.

**This study aimed** to assess the effects of non-toxic  $\text{Cu}^{2+}$  levels on death of cultivated cerebellar granule neurons induced by lipopolysaccharide (LPS; *in vitro* inflammation model) or azide sodium ( $\text{NaN}_3$ ; cytochrome c oxidase inhibitor).

## Materials and methods

In our experiments, we used 7-day to 8-day cultures of the cerebellum from 8-day-old rats obtained by enzymatic and mechanical dissociation: 15 minutes at 36.5°C in trypsin (0.05%) and EDTA (0.02%) solution in phosphate buffer (Gibco Life Technologies) followed by stepwise pipetting in the medium [10]. The cultures were cultured in 96-well plastic plates (Eppendorf) coated with poly-lysine (Sigma). The culture medium contained 90% minimum essential medium with Earle's salts (Gibco), 10% fetal bovine serum (HyClone), 2 mM glutamine (glutaMAX, Gibco), 25  $\mu\text{M}$  KCl, and 10 mM HEPES buffer pH 7.2 to 7.4 (VWR Life Science). To each plate well, 0.1 mL of cell suspension was added to obtain a final cell density of 3 to 5  $\times 10^3$  cells/ $\text{mm}^2$ . The cultures were developed in a  $\text{CO}_2$ -incubator at 36.5°C and relative humidity 98%.

On day 7 to 8, copper (II) chloride (0.5 to 5.0  $\mu\text{M}$ , Sigma) with LPS (10  $\mu\text{g/mL}$ , Sigma) or  $\text{NaN}_3$  (250  $\mu\text{M}$ ) was added to the culture medium with 7-day rat cerebellar cells *in vitro* for 24 hours.

After the experiment, the cultures were fixed in ethanol + formaldehyde + acetic acid mixture (7 : 2 : 1) and stained with trypan blue. The cultures were photographed with an Olympus CKX41 inverted microscope

or an EVOS M7000 imaging system (Termo Fisher Scientific) at  $\times 40$  objective magnification. Percentage of survived neurons was evaluated by counting morphologically intact cultured granule neurons in 5 consecutive field views. Survival in test cultures was expressed in per cent vs. control.

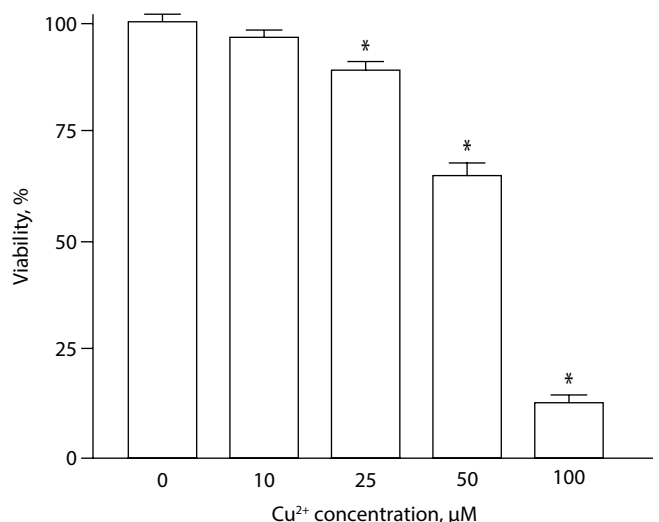
The level of nitric oxide (NO) was determined by Griess assay, which is based on the formation of diazo compounds that react with alpha-naphthylamine to give a red solution. Photometry was performed with a microplate reader (SpectraMax M2, Molecular Devices) at 540 nm.

Data were statistically processed with Statistica v. 13.3 (StatSoft Inc.) and one-way ANOVA with Newman–Keuls post hoc test or t-test. Between-group differences were considered statistically significant if  $p < 0.05$ . Results were presented as mean  $\pm$  standard error of mean ( $M \pm SEM$ ).

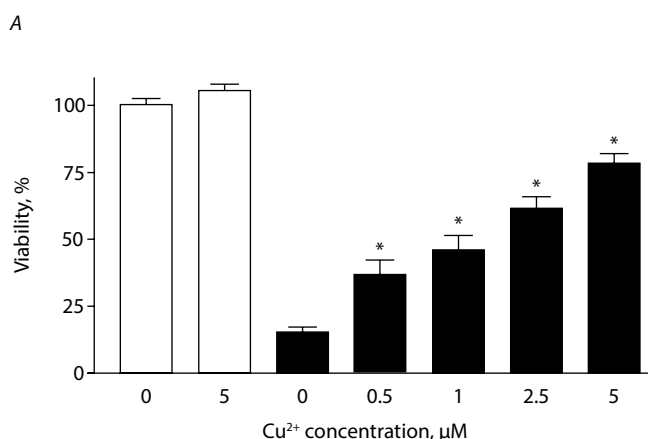
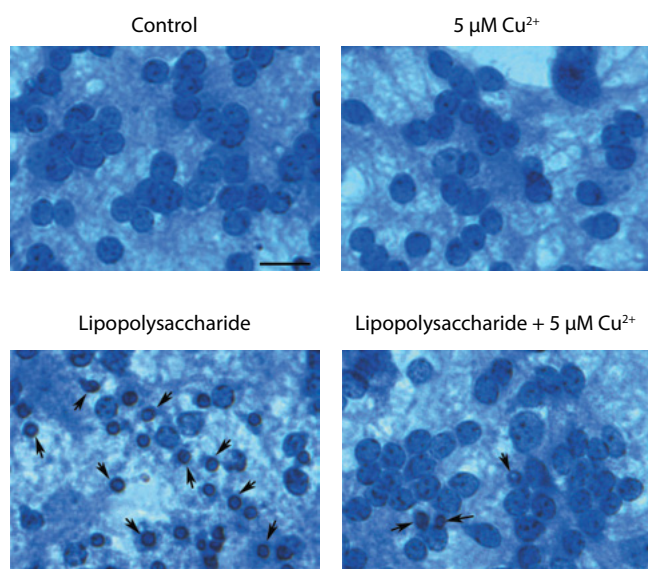
All procedures performed in the experiments involving animals complied with the ethical standards approved by the Russian regulations, the principles of the Basel Declaration, and Recommendations of the Local Ethical Committee of Research Center of Neurology (Protocol 5-5/22 of 1 June 2022).

## Results

$\text{Cu}^{2+}$  toxicity in cultured cells was seen with concentrations of at least 25  $\mu\text{M}$ . With further increase in  $\text{Cu}^{2+}$  concentrations, neuron survival decreased in a dose-dependent manner (Fig. 1). Added to the culture medium, LPS or  $\text{NaN}_3$  decreased neuron survival



**Fig. 1. Effects of different copper ion levels on survival of cultured rat cerebellar granule neurons.**  
\* $p < 0.05$  vs. control (0  $\mu\text{M}$ ).



**Fig. 2. Copper ions reduced LPS toxicity in cultured rat cerebellar granule neurons.**

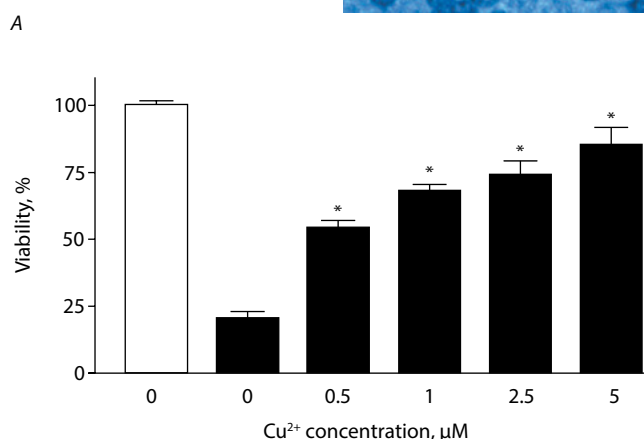
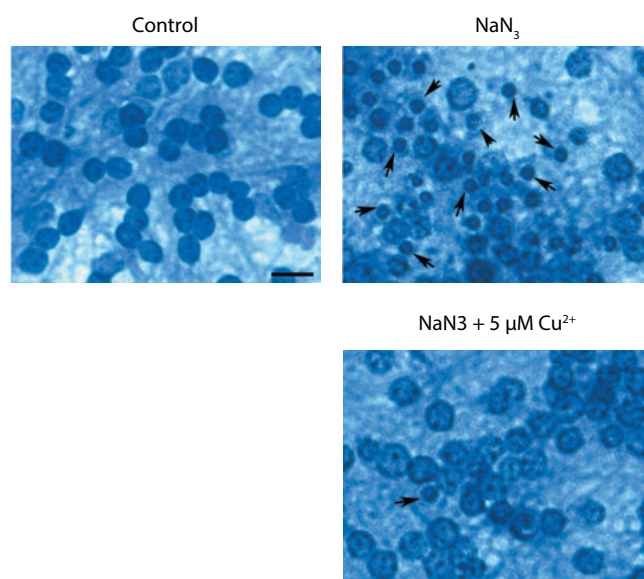
A: fixed cultures stained with trypan blue. Dead neuron nuclei are shown with arrows. Scale 15 μm.

B: quantitative data obtained by counting morphologically intact neurons without (white bars) and with LPS (black bars).

\* $p < 0.05$  compared to 0 μM Cu<sup>2+</sup> with LPS.

to  $15 \pm 2\%$  (Fig. 2) or  $20.0 \pm 2.5\%$  (Fig. 3) vs. control, respectively. If neurons were treated with the toxins in the presence of non-toxic copper ion levels, neuron survival increased in a dose-dependent manner.

Cu<sup>2+</sup> 5 μM improved neuron survival to  $78 \pm 4\%$  in the experiment with LPS (Fig. 2) or to  $86 \pm 6\%$  in the experiment with NaN<sub>3</sub> (Fig. 3). The concentration of nitrites in the control culture medium was  $2.0 \pm 0.2$  μM. Added to the cell cultures, LPS increased the concentration of nitrites to  $8.5 \pm 0.5$  μM (Fig. 4). Copper chloride 5 μM did not have any significant effects on nitrite accumulation in the culture medium treated with LPS (Fig. 4).



**Fig. 3. Copper ions reduce NaN<sub>3</sub> toxicity in cultured rat cerebellar granule neurons.**

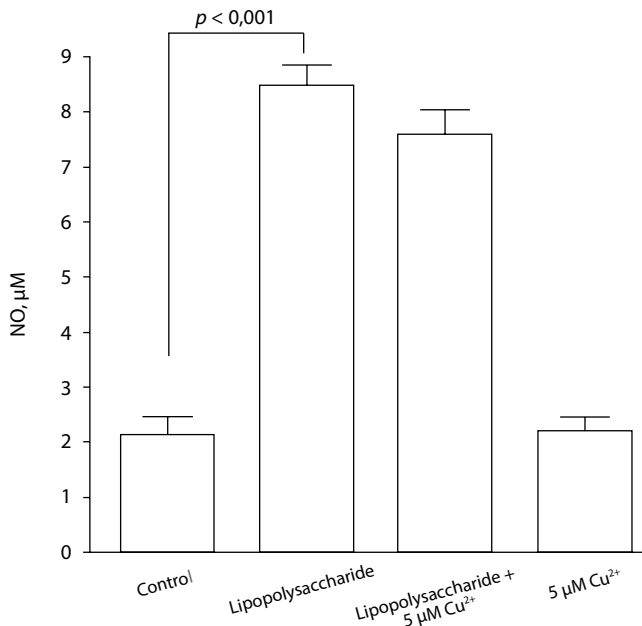
A: fixed cultures stained with trypan blue. Dead neuron nuclei are shown with arrows. Scale 15 μm.

B: quantitative data obtained by counting morphologically intact neurons without (white bars) and with NaN<sub>3</sub> (black bars).

\* $p < 0.05$  compared to 0 μM Cu<sup>2+</sup> with NaN<sub>3</sub>.

## Discussion

Imbalances of several metal ions, especially zinc and copper, are thought to play an important role in the pathogenesis of many neurodegenerative disorders, including multiple system atrophy, amyotrophic lateral sclerosis, Creutzfeldt–Jakob disease, Wilson disease, Alzheimer’s disease, and Parkinson’s disease [1, 11, 12]. Normally, copper ions are structural elements of many proteins, including ceruloplasmin, a blood plasma protein that is involved in the mechanisms of various pro-oxidant and antioxidant reactions. Copper is necessary for functioning of the antioxidant cell system because it is contained in the superoxide dismutase molecule. Cu



**Fig. 4. The levels of nitrites (NO) in the culture medium of rat cerebellar granule neurons.**

The addition of LPS (10 µg/ml, 24 h) causes an increase in nitrites in the culture medium. Cu<sup>2+</sup> (5 µM) have no significant effect on the accumulation of nitrite in the culture medium under LPS action.

(II) derivatives are effective anti-inflammatory agents [13, 14], and Cu-binding peptides showed anti-inflammatory effects in primary microglia cultures [15].

NO is a key inflammation mediator. Glia cells with inflammatory activation, which is seen in most CNS disorders, were previously shown to be capable of exerting neuronal toxicity, which was prevented by inhibitors of inducible NO synthase [16]. Excessive formation of NO or reactive NO species, such as peroxynitrite, impairs mitochondrial functioning and eventually affects neuronal cell metabolism and survival [17, 18]. Besides its multiple regulatory functions, NO was found to modulate cell respiration by irreversibly inhibiting the cytochrome c oxidase activity [19, 20].

In our study we showed that LPS, which was added to the culture medium with neuroglia cultures, reduced survival of cultured rat cerebellar granule neurons and was associated with nitrite accumulation in the culture

medium due to NO production. Added to the culture medium, non-toxic concentrations of Cu<sup>2+</sup> significantly reduced LPS-induced cell death. NO is known to act as a ligand for copper atoms and cause a redox reaction with the metal after its binding. Furthermore, NO possesses an unpaired electron, which can couple with the unpaired electron on Cu<sup>2+</sup> [21]. In our experiments, copper did not show any significant effects on nitrite accumulation in the culture medium treated with LPS. Moreover, NO can inhibit mitochondrial respiration mainly by competitively inhibiting oxygen binding by Cu<sup>2+</sup>-containing cytochrome c oxidase (Complex IV) [22] and direct interaction of Cu<sup>2+</sup> with tricarboxylic acid cycle enzymes [23]. Our experiments demonstrated that copper ions protected neurons against the toxicity induced by NaN<sub>3</sub>, which inhibits Complex IV of the electron transfer chain in the mitochondria.

Our data correlate with previous results that showed that pretreatment with CuSO<sub>4</sub> prevented inhibition of mitochondrial complexes I, II, IV, V and Cu/Zn-superoxide dismutase induced by 1-methyl-4-phenylpyridine (MPP<sup>+</sup>) in the rat striatum [24]. In this neurodegeneration model, CuSO<sub>4</sub> also reduced the MPP<sup>+</sup>-induced increase in the enzymatic activity levels of caspases 8, 9 and 3, decreased apoptotic cell damage [25], and prevented the hypokinetic state in MPP<sup>+</sup>-treated mice [26]. In mice, a copper-chelator led to reduced activity of complex IV in neurons and dropped activity of the anti-oxidant system in the brain tissue [27, 28]. Based on the above data, we can assume that the protective effect of copper ions in inhibiting of electron transport chain complexes may be associated with a direct effect on copper-dependent proteins or an indirect effect on apoptotic pathway proteins.

## Conclusion

We showed that Cu<sup>2+</sup> protected neurons against the toxicity induced by LPS, an inflammation inductor, or NaN<sub>3</sub>, a cytochrome c oxidase inhibitor. This protection is likely to be associated rather with Cu<sup>2+</sup> interaction with Complex IV of the electron transfer chain in the mitochondria than with inhibition of NO production. Effects of Cu<sup>2+</sup> on apoptosis pathway proteins also cannot be ruled out.

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